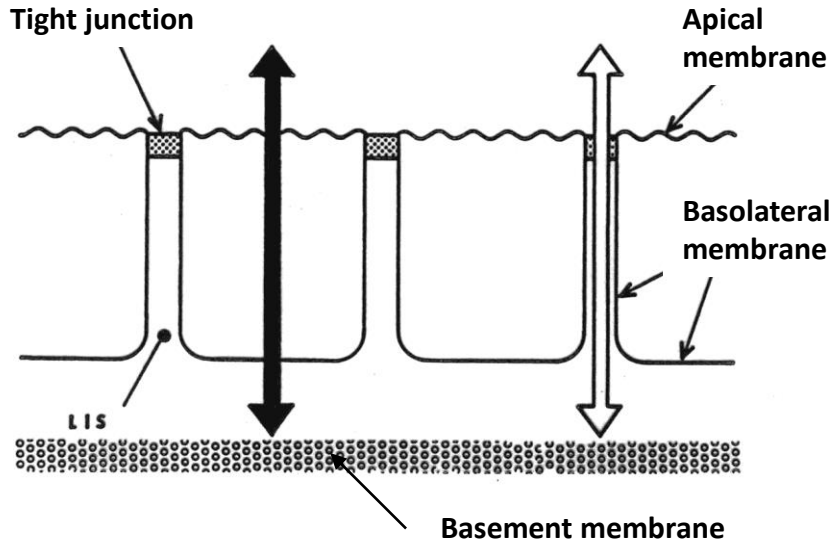


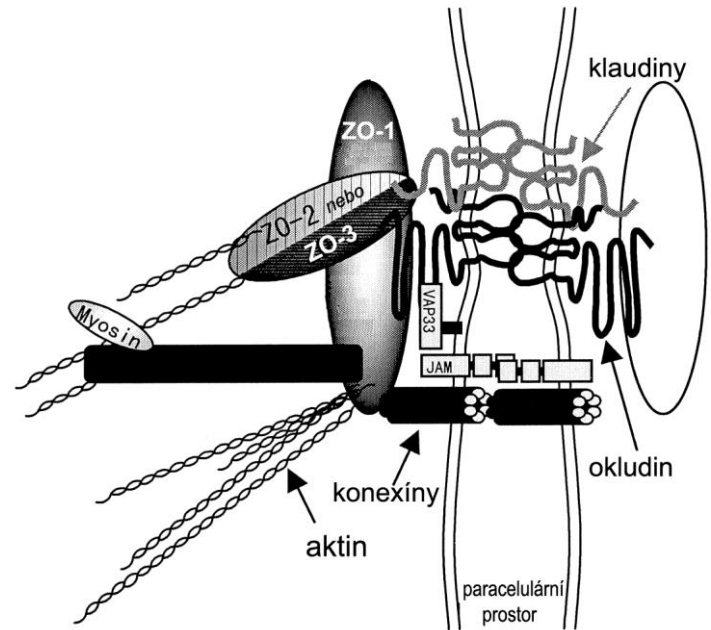
Model of the epithelium and its principal compartments



↔ Transcellular flux

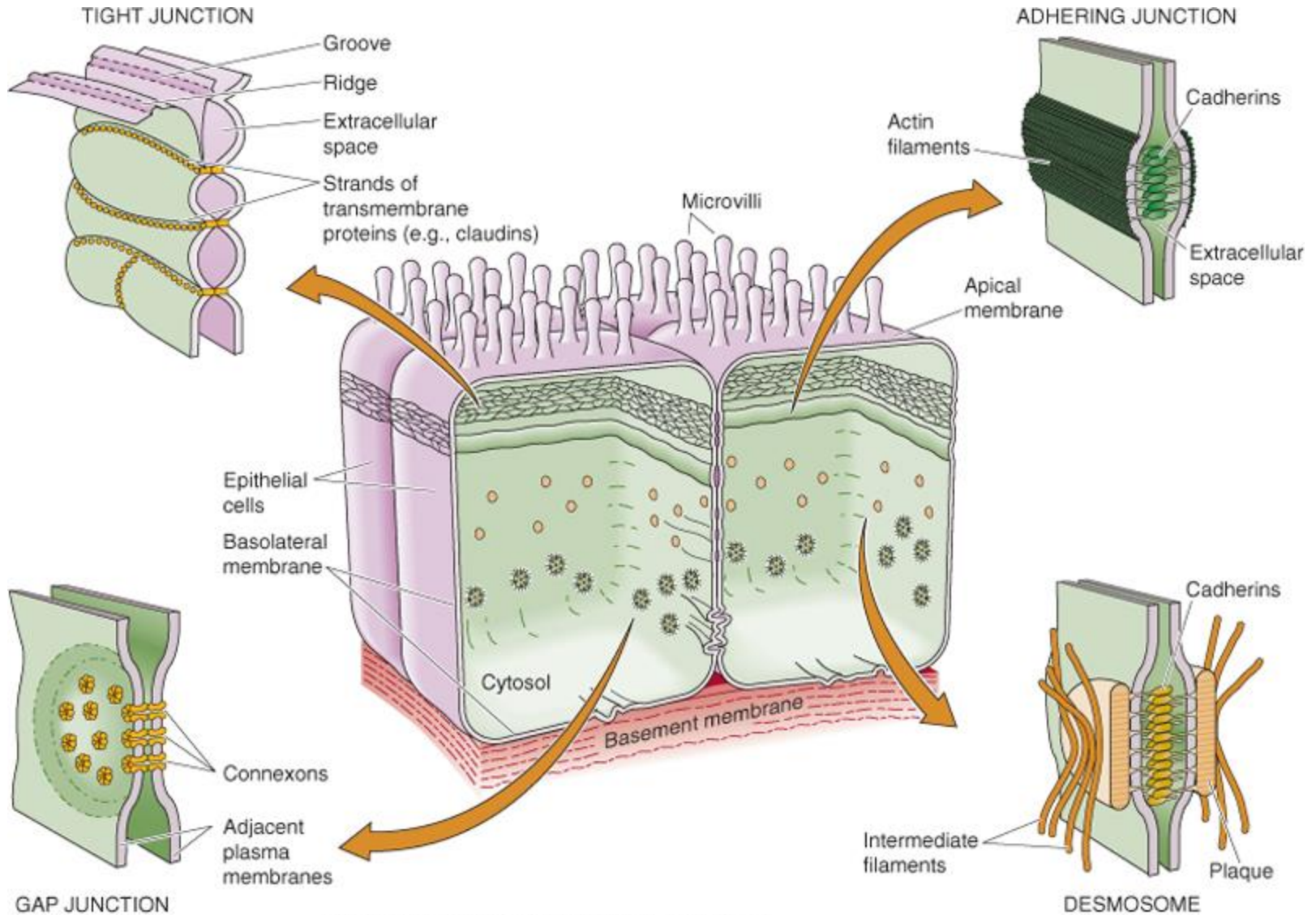
↔ Paracellular flux

LIS - Lateral intercellular space

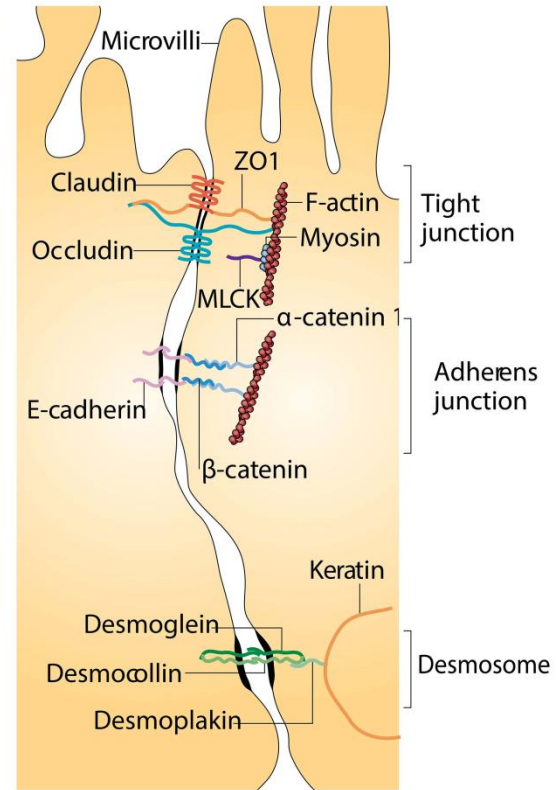
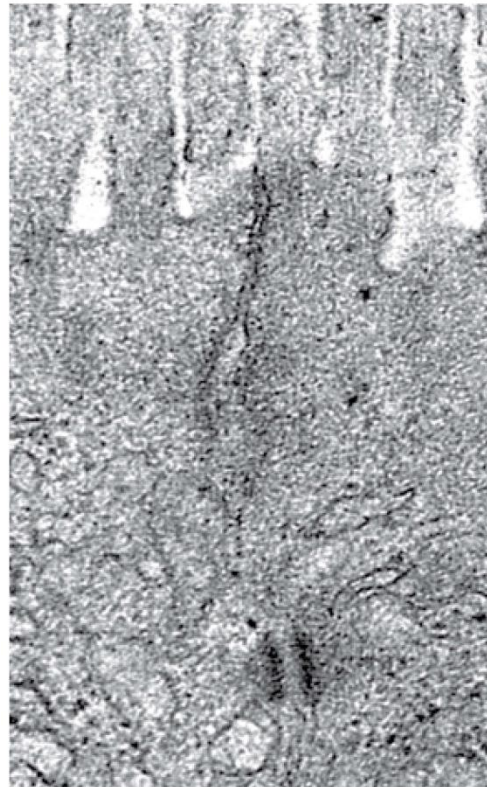


Tight junction

Epithelial cell and its junction complexes



Epithelial cell and its junction complexes



Interaction of epithelial cells with basement membrane

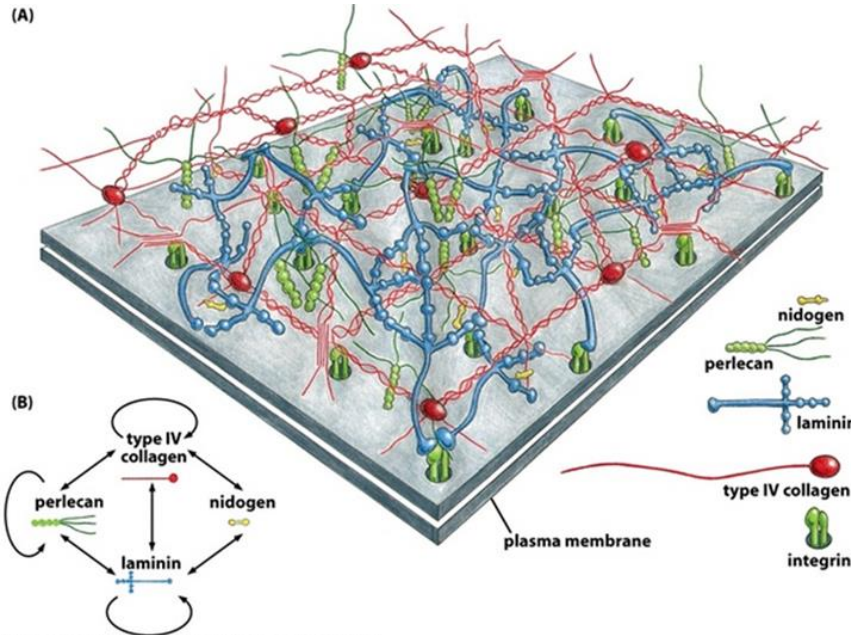
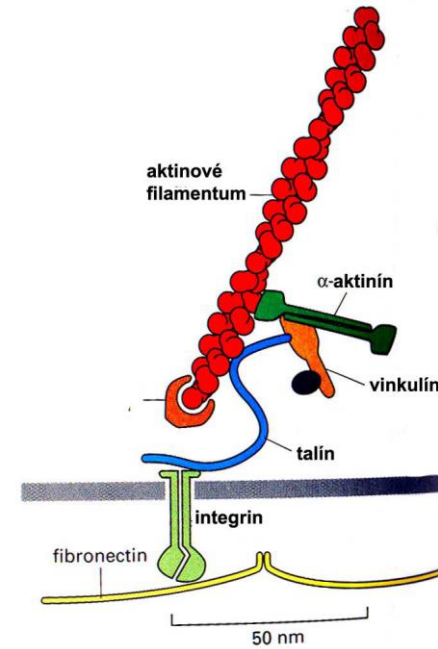
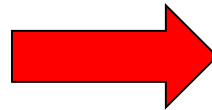


Figure 19-43 Molecular Biology of the Cell 5/e (© Garland Science 2008)

Principal proteins of the basement membrane

Epithelial cell is attached to the basement membrane via integrins (heterodimer receptors)



Transport of solutes across epithelium

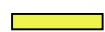
$$J_i = \underbrace{(1-\sigma_t) \cdot C_i \cdot J_V}_{\text{„solvent drag“}} + \underbrace{P_i \cdot (\Delta c_i + z_i \cdot F / \Delta \varphi \cdot C_i)}_{\text{electrodifusion}} + \underbrace{J_i^{\text{akt}}}_{\text{active transport}}$$



„solvent drag“



electrodifusion



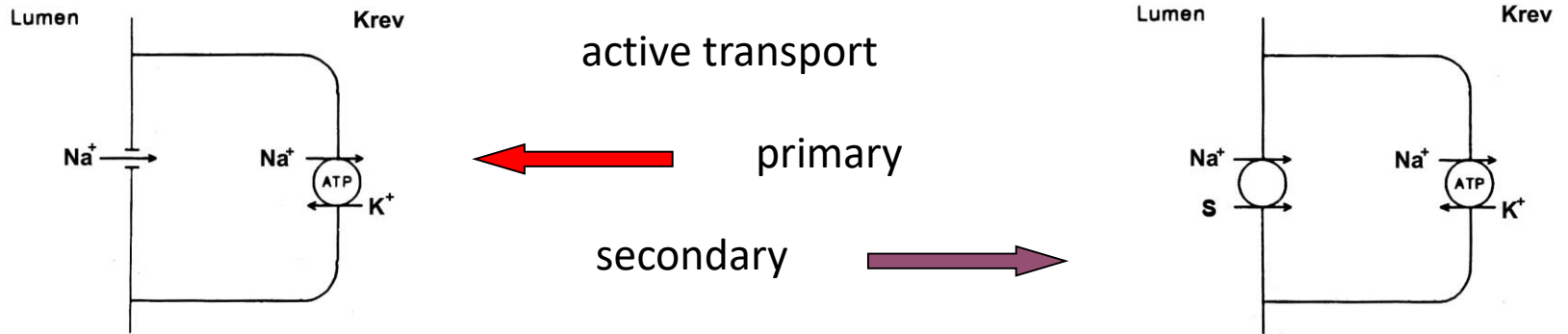
active transport

- J_i Flux of transported molecule/ion
- σ_t Staverman reflection coefficient
- C_i Intraepithelial concentration of the solute i
- J_V Volume flow
- P_i Permeability coefficient
- Δc_i Concentration difference mucosa-serosa
- z_i Valency
- F Faraday number
- $\Delta \varphi$ Transepithelial voltage

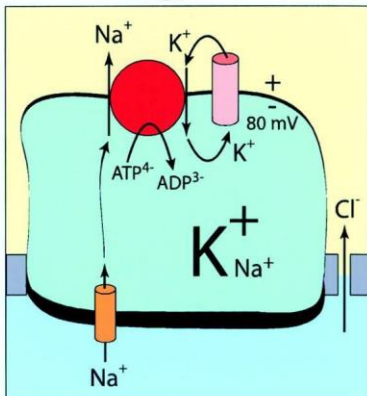
NaCl transport in renal proximal tubule

	J_i			
Na	100 %	29 %	32 %	38 %
Cl	100 %	49 %	51 %	0 %

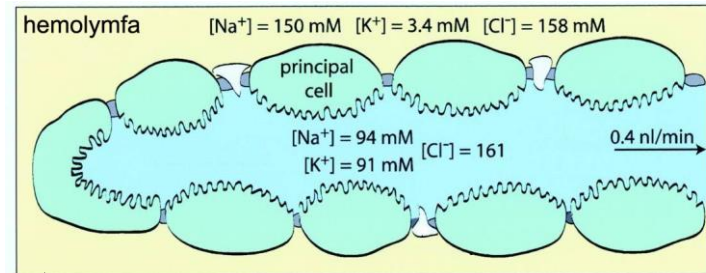
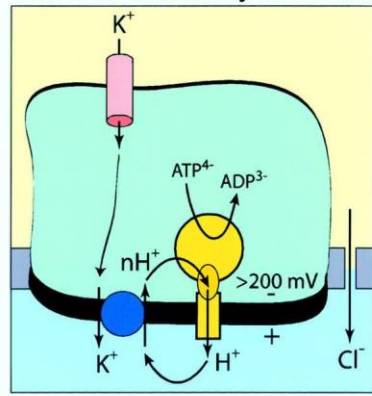
Transport of solutes



Ussingův model



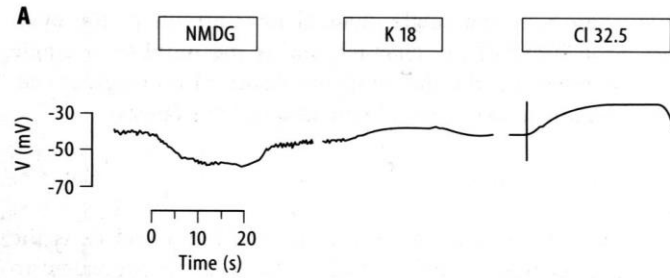
Wieczorek/Harveyův model



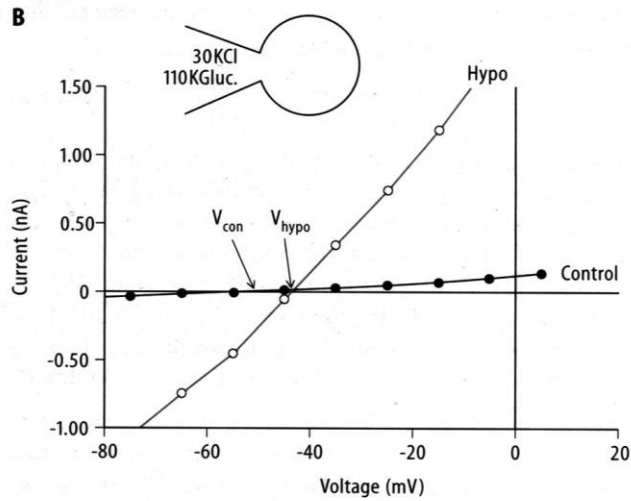
Malpighian tubule (Insects)

Frog skin, urinary bladder, colon, renal collecting tubule, etc.

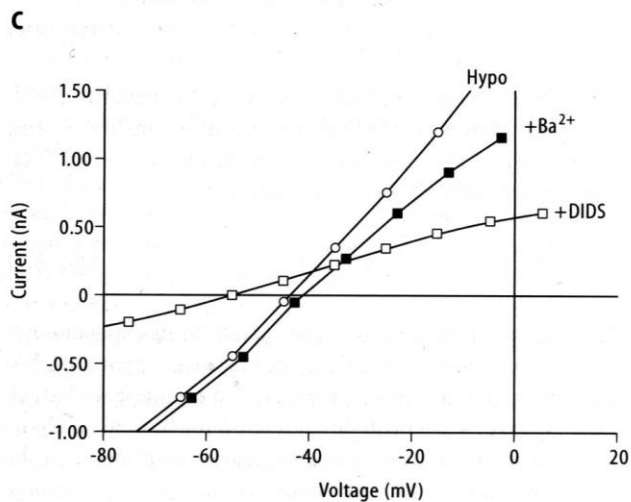
Conductance of epithelial cells



Ion conductances of plasma membrane, membrane potentials

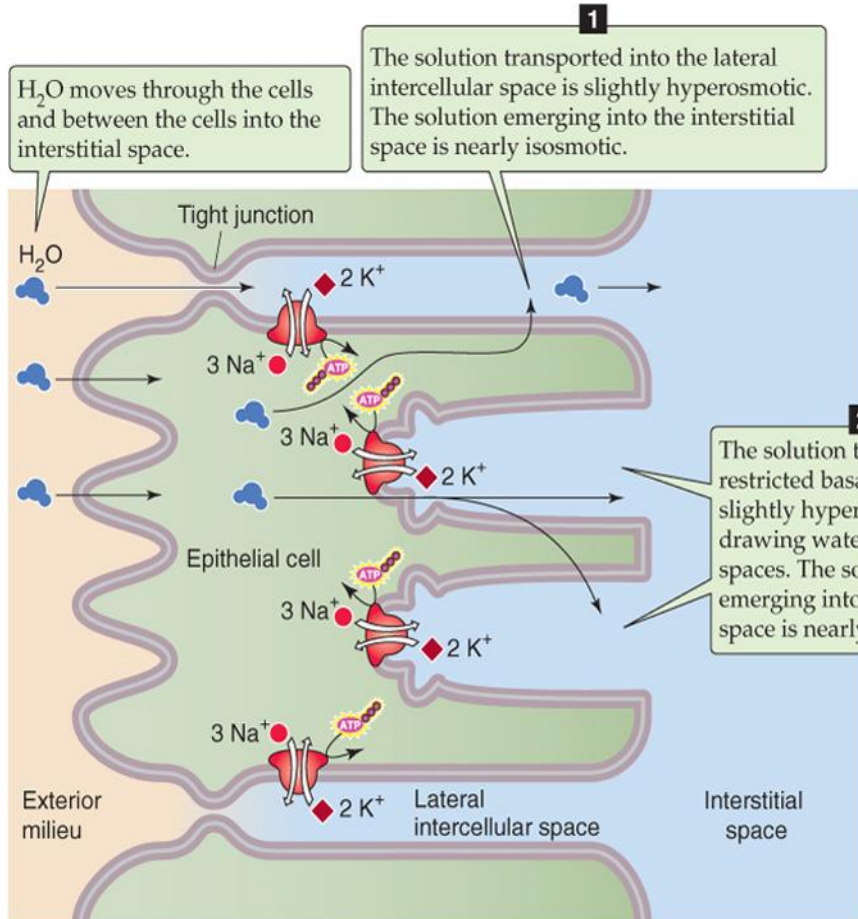


Effect of hypotonic stress on membrane conductance



Barium and DIDS sensitive conductance is induced by hypotonic stress

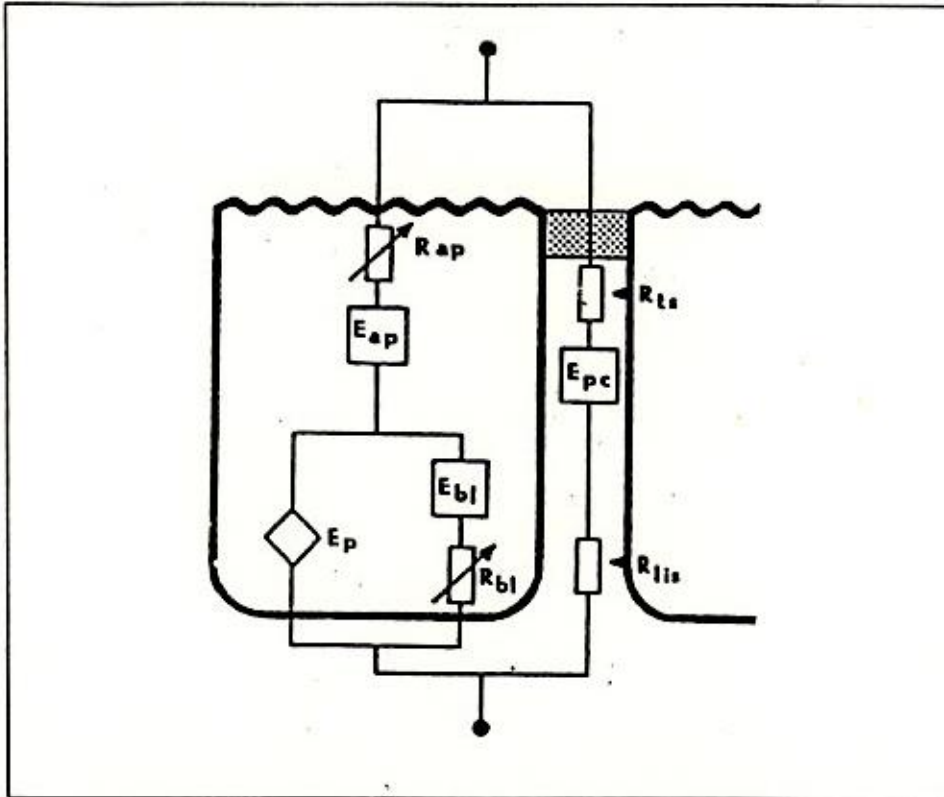
Transport of solutes and water



$$J_v = L_p(\Delta p - R.T.\Delta c_1 - \sigma_t. R.T.\Delta c_2)$$

- J_v Volume flow
- L_p Hydraulic conductivity
- Δp Hydrostatic pressure difference
- R Gas constant
- T Absolute temperature
- Δc_1 Concentration difference mucosa-serosa
- Δc_2 Concentration difference mucosa-serosa
- σ_t Staverman reflection coefficient

Equivalent electrical circuit for the epithelium



R_{ap} = parallel resistance (1/conductance) for individual ion species that move across the apical membrane

E_{ap} = parallel electromotive forces equal to Nernst equilibrium potentials of individual ion species that move across the apical membrane

R_{bl}, E_{bl} = the same for the basolateral membrane

E_p = electromotive force of the electrogenic ion pump of the basolateral membrane (Na,K-ATPase)

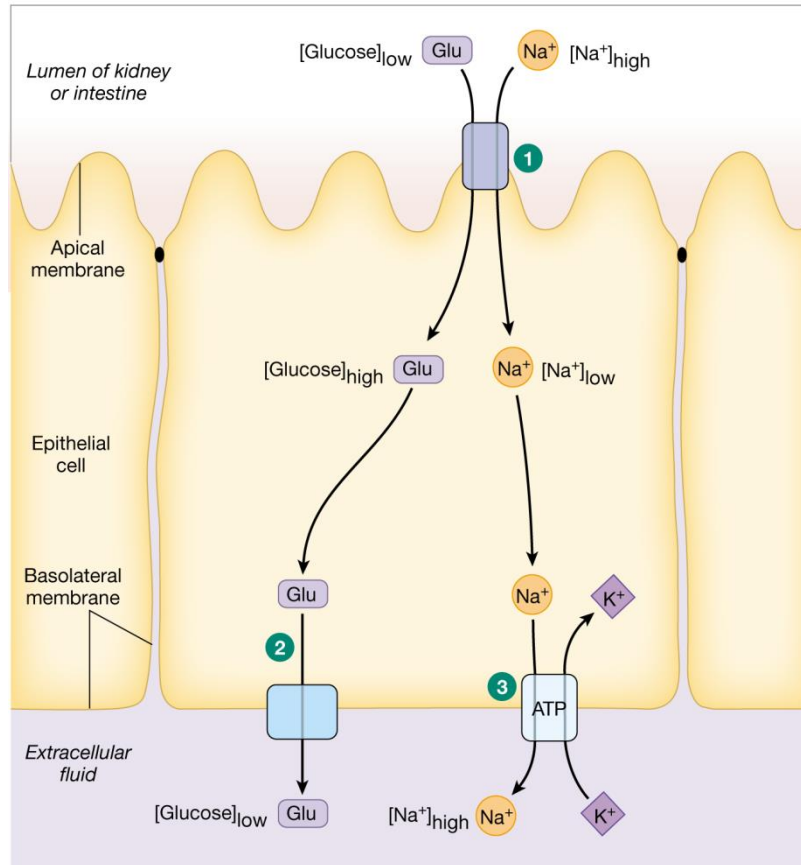
R_{ts} = resistance of the tight junction

E_{pc} = electromotive force corresponding to the diffusion potential of the tight junction

R_{lis} = resistance of the lateral intercellular space

Coupling of apical and basolateral transport

Absorbing glucose from intestinal or kidney tubule lumen involves indirect (secondary) active transport of glucose across the apical membrane and glucose diffusion across the basolateral membrane.



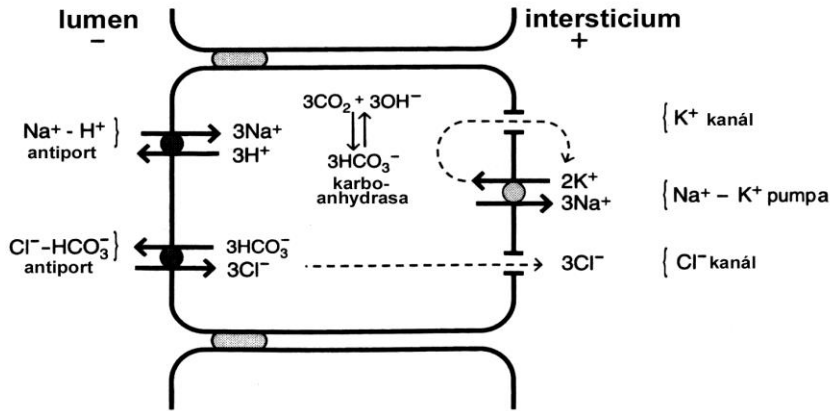
1 Na⁺-glucose symporter
brings glucose into cell against its gradient using energy stored in the Na⁺ concentration gradient.

2 GLUT transporter
transfers glucose to ECF by facilitated diffusion.

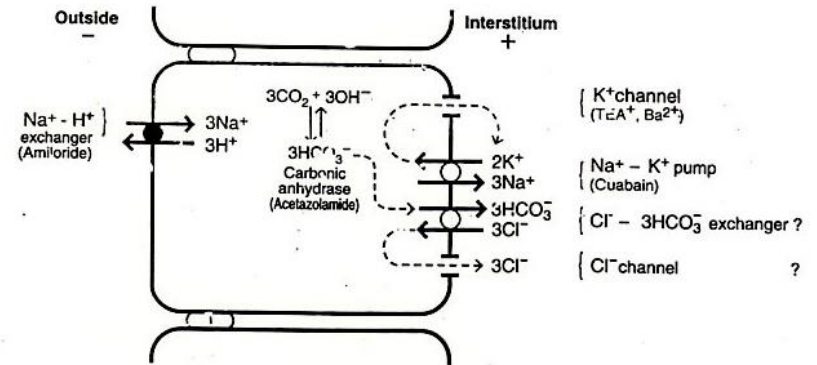
3 Na⁺-K⁺-ATPase pumps Na⁺ out of the cell, keeping ICF Na⁺ concentration low.

- Secondary active transport has to be coupled with primary active transport (1) and (3)

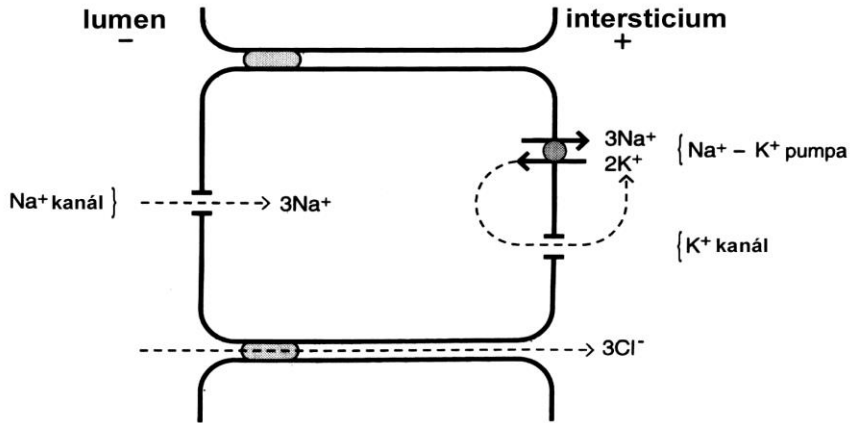
Mechanisms of NaCl (NaHCO₃) absorption



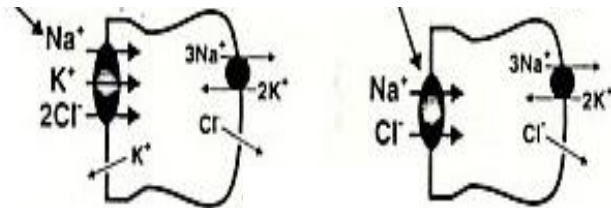
Transcellular NaCl transport



Transcellular NaHCO₃ transport



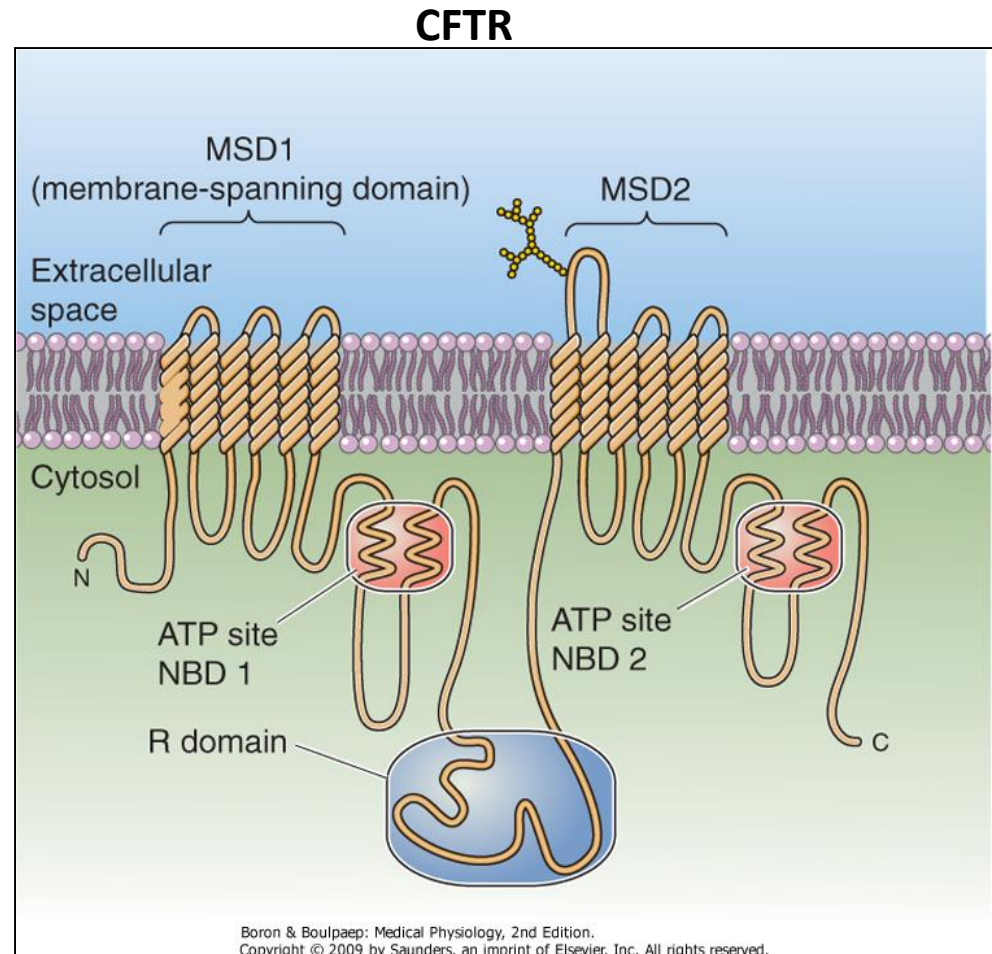
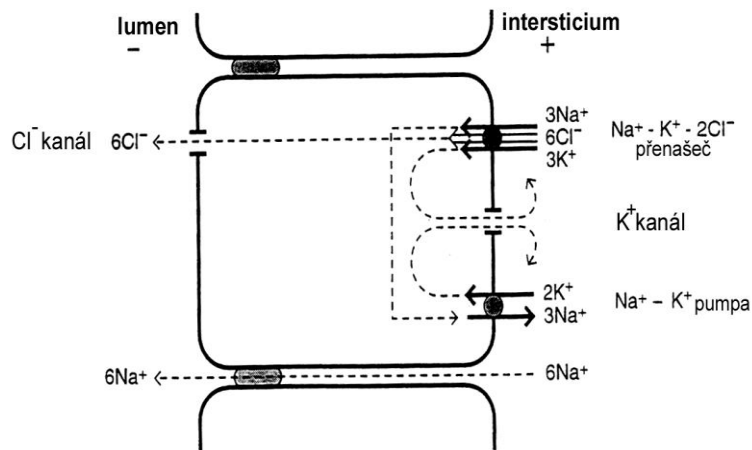
Transcellular transport of Na⁺ and paracellular transport of Cl⁻



Transcellular NaCl transport

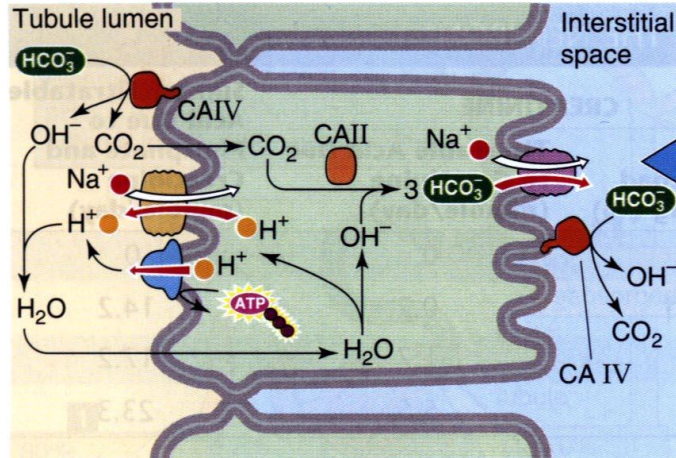
NaCl and water secretion

Transcellular transport of Cl^- via NKCC cotransporter on the basolateral membrane and chloride channel CFTR on the apical membrane. The negative charges that move across the cell from the interstitium to lumen generate a lumen-negative voltage that can drive passive Na^+ secretion across the paracellular pathway (tight junctions). The driving force is the secondary active transport of Cl^- across the basolateral membrane.

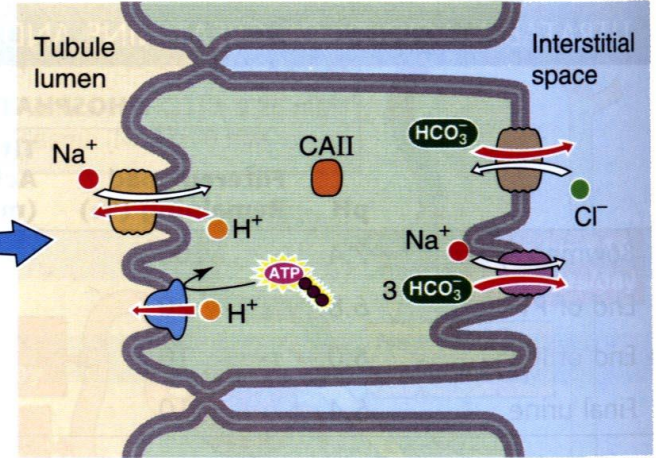


Acid-base transport – luminal acidification

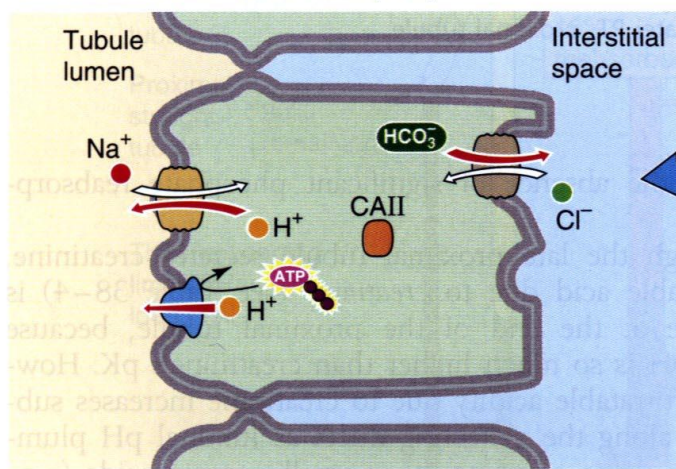
A EARLY PROXIMAL CONVOLUTED TUBULE (S1)



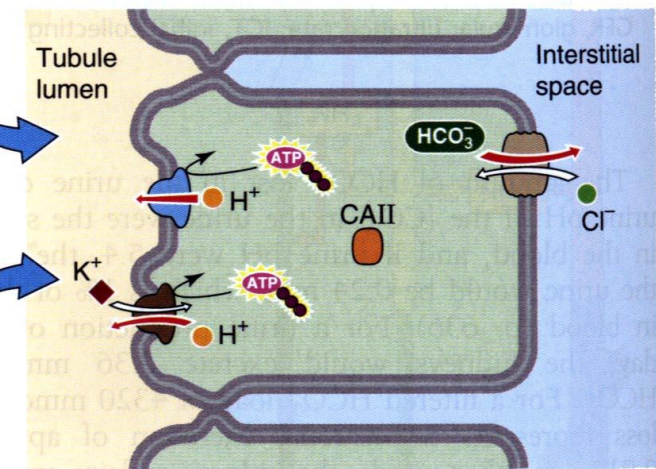
B LATE PROXIMAL STRAIGHT TUBULE (S3)



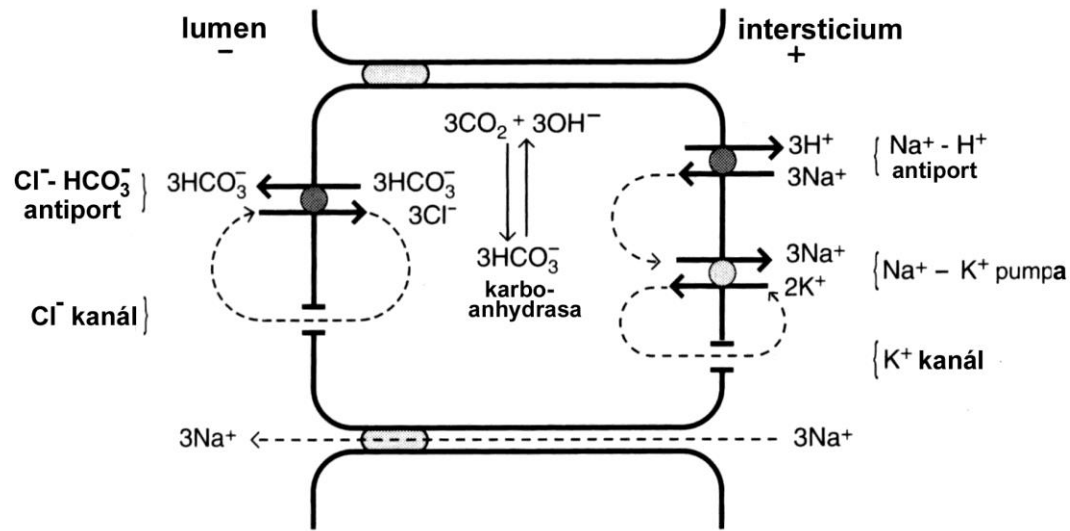
C THICK ASCENDING LIMB (TAL)



D α INTERCALATED AND MEDULLARY COLLECTING-DUCT CELLS

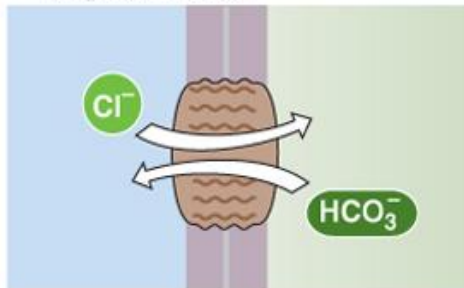


Acid-base transport – luminal alkalization

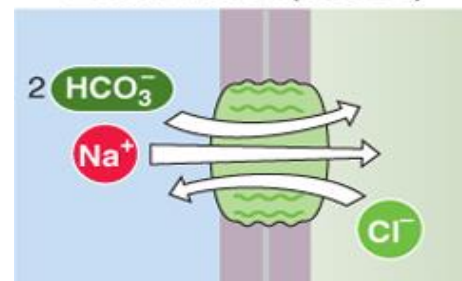


Exchangers

D $\text{Cl}^- - \text{HCO}_3^-$ EXCHANGER (e.g., AE, DRA)

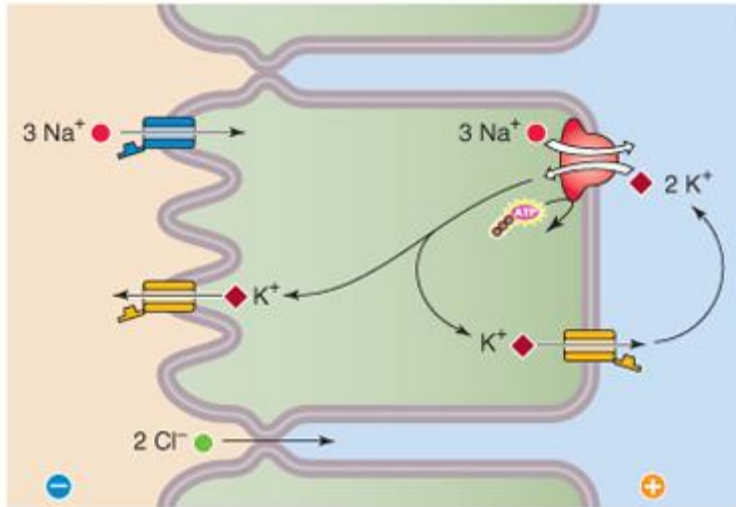


C Na-DRIVEN $\text{Cl}^- - \text{HCO}_3^-$ EXCHANGER (NDCBE)

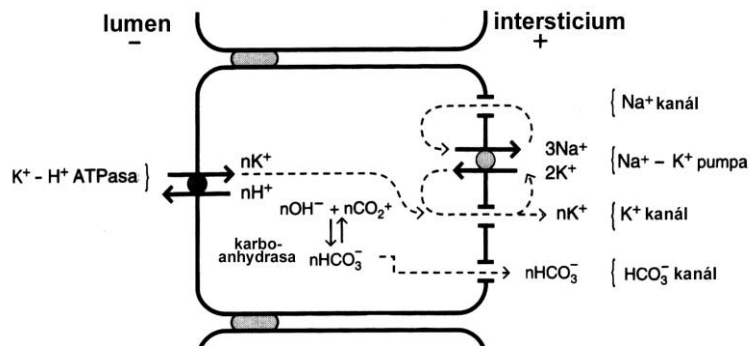


Potassium transport

B K⁺ SECRETION

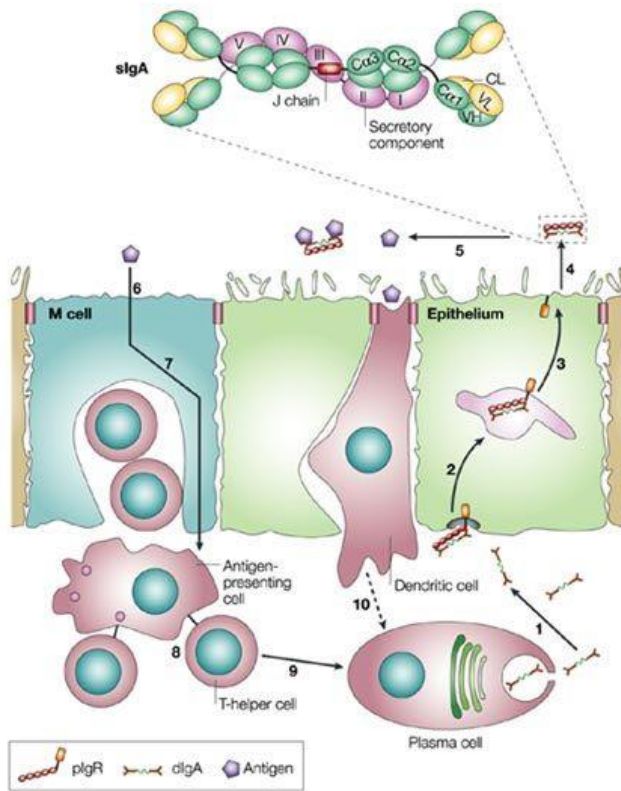


Transcellular secretion: K⁺ is taken up by Na-K pump across the basolateral membrane to be secreted across the apical membrane via potassium channels.



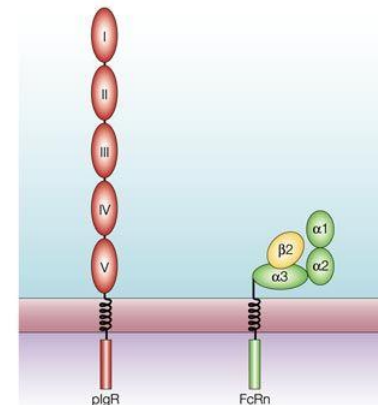
1. Paracellular absorption through tight junctions.
2. Transcellular primarily active absorption driven by H-K-pump.

Immunoglobulin transport across polarized epithelial cells



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- Receptor mediated transcytosis
- Transport of IgA and IgG is mediated through the polymeric immunoglobulin receptor pIgR
- pIgR is proteolytically cleaved in the cell or at the apical surface
- The cleaved extracellular domain of the receptor = secretory component (SC)
- Transport of IgG is mediated through FcRn receptor



Nature Reviews | Molecular Cell Biology

Control of epithelial transport

1. **Cell control** – „crosstalk“ between apical and basolateral membrane – protection against the increasing concentration of osmolytes in the cytoplasm
2. **Tissue control** – e.g.. enteric nervous system,
3. **Organism control** – e.g. hormones calcitriol, aldosterone, vasopressin

Hormone regulatory pathways – endocrine, paracrine, autocrine, intracrine

Nervous regulatory pathways – symphatetic, parasympathetic and enteric nervous systems

Immune regulatory pathways – mucosal immune system

